Packing List

* Secchi disk
* PVC integrated sampler
* Bucket
* Clear plastic pitcher for collecting water from integrated sampler
* 1 L bottles
  + Littoral only: 6 acid-washed 1 L Nalgene bottles (includes 1 extra)
  + Littoral + Pelagic: 12 acid-washed 1 L Nalgene botles
* Hydrolab sonde with PCY probe (NOT the one with turbidity)
* Rope and cord for Hydrolab sonde
* Both surveyors
* 8 backup C batteries for Hydrolab sonde
* White plastic sampling cage for hydrolab sonde
* cooler with ice (x 2 for littoral + pelagic)
* Pencils, sharpie markers for labeling bottles
* data sheets in binder (plus special OWRB data sheets on clipboard
* 63 µm zooplankton net + rope + spare binder clips (for pelagic, bring a second net)
* Zoop bottles
  + Littoral only: 11 x 250 ml zooplankton bottles with ethanol added for preserving tows, marked with max fill line (includes 1 extra)
  + Littoral + Pelagic: 22 x 250 ml zooplankton bottles with ethanol added
* DI squirt bottle
* Kestrel
* Printed cryo-labels for 2 ml tubes
* 1 L Nalgene filled with DI water (spare water for rinsing zoop nets)- bring extra for pelagic

Before going out

1. Check zoop net for snags in netting, detritus, or cracked latex tubing
2. Calibrate Hydrolab sonde and charge surveyors
3. Acid wash sample bottles
   1. For littoral only: 6 x 1 L Nalgene bottles
   2. For littoral + pelagic: 12 x 1 L Nalgene
4. Collect ice in coolers
   1. For littoral only: One cooler for 6 x 1 L nalgenes + 11 zoop bottles
   2. For littoral + pelagic: Two coolers that each fit 6 x 1 L Nalgenes + 11 zoop bottles
5. Mark 250 ml zooplankton bottles with 125 ml max fill line, measure out 90 ml 95% ethanol and 1.25 ml glycerin into each bottle (10 total). Pre-label bottles and caps with sampling site and date.
   1. For littoral only: 11 bottles
   2. For littoral + pelagic: 22 bottles
6. Pre-label 1L nalgene bottles and caps with sampling site and date. One bottle per site.
7. Autoclave 40 Pall 0.2 µm supor filters for DNA samples and dry overnight in drying oven.