Packing List

* Secchi disk with rope
* PVC integrated sampler with rope
* Blue bucket
* White bucket with black sharpie marks inside
* Clear plastic pitcher for collecting water from integrated sampler
* 1 L bottles
  + Littoral only: 6 acid-washed 1 L Nalgene bottles (includes 1 extra)
  + Littoral + Pelagic: 12 acid-washed 1 L Nalgene botles
* Hydrolab sonde with PCY probe (NOT the one with turbidity)
* Rope and short cord for Hydrolab sonde. Long cord may be needed for deep sites.
* Both surveyors
* 8 backup C batteries for Hydrolab sonde
* White plastic sampling cage for hydrolab sonde
* cooler with ice (x 2 for littoral + pelagic)
* Pencils, sharpie markers for labeling bottles
* data sheets in binder (plus special OWRB data sheets on clipboard)
* 63 µm zooplankton net + rope + spare binder clips (for pelagic, bring a second net) + zooplankton cup
* Zoop bottles
  + Littoral only: 11 x 250 ml zooplankton bottles with ethanol added for preserving tows, marked with max fill line (includes 1 extra)
  + Littoral + Pelagic: 22 x 250 ml zooplankton bottles with ethanol added
* DI squirt bottle (x 2 for pelagic)
* Kestrel
* 2x 1 L Nalgene filled with DI water (spare water for rinsing zoop nets)- x 2 for pelagic
* FLAME
* FLAME computer

Before going out

1. Check zoop net for snags in netting, detritus, or cracked latex tubing
2. Calibrate Hydrolab sonde and charge surveyors
3. Acid wash 1 L Nalgene sample bottles if not already washed (number to be determined by lake: 1 x number of lakes)
4. Pre-label 1L nalgene bottles and caps with sampling site and date. One bottle per site.
5. Mark 250 ml zooplankton bottles with 125 ml max fill line, measure out 90 ml 95% ethanol and 1.25 ml glycerin into each bottle (10 total). Pre-label bottles and caps with sampling site and date. Number of bottles to be determined by site (2 x number of sites).
6. Print labels for GF/F and 0.2 µm filters.
7. Make sure there are enough field data sheets in binder. Print if needed.
8. Make sure there are enough field data sheets for OWRB (half sheets). Print if needed.
9. Autoclave 40 Pall 0.2 µm supor filters for DNA samples and dry overnight in drying oven.
10. Make sure there are enough 10 ml tips (clean + autoclaved).
11. Pre-label 50 ml tubes: 5 per site + 1 blank (2 whole water, 2 GF/F filtrate for PELL, 1 whole water for USGS, 1 DI water for USGS) – see tube labeling protocol.
12. Pre-label zooplankton vials (tops + paper slips).
13. Check for Lugol’s tubes and prep if more are needed. We need one per site- 15 ml opaque centrifuge tube with 100 µl Lugol’s solution.
14. Day of sampling: collect ice in coolers